

THEILERIOSIS INFECTION IN ANIMALS (ARTICLE REVIEW)

1 Russul Wassit Kadhum,

2 Zahraa Ali Faieq,

3 Zainab Abbas Jasim Al-Maliki

1, 2, 3 Department of Biology, College of Sciences, University of Wasit, Iraq

Corresponding author: russul.wassit@uowasit.edu.iq

Abstract

Theileria-genus blood parasites are the cause of the protozoan disease theileriosis (1). Apical complexes, a distinct set of unique organelles, are used to identify parasites (2). One of the most prevalent tick-borne illnesses is the parasite *Thoilor*, which has been linked to cases in a variety of ruminants, including goats, cattle, and sheep. Theileria are obligate intracellular parasites that are spread by ticks and pose a significant threat to livestock in tropical and subtropical areas of the world. The primary vector of infection for Theileria, *Hylomma dromodarii* ticks, infested those animals, suggesting a function for the parasite's development with a range of forms and stages inside this vector. (3). In general, all *Thoilor* species have a similar life cycle. It acknowledged two hosts—vertebrate hosts and tick vectors—and allowed two forms: lymphatic and erythrocyte (4).

Introduction

Life cycle of Theileria parasite

Theileria parasites have a complicated and dynamic life cycle they are dispersed as vector ticks of proliferation phases and have a generic apicomplexan with multiple differentiation steps. (5). The life cycle of the parasite consists of at least three stages: schizogony and merogony, which are forms of asexual reproduction in the vertebrate host, and sporogony, which is a kind of sexual reproduction in the tick vector. During tick feeding, Theileria sporozoites infiltrate their vertebrate host and swiftly arrive at mononuclear leukocytes, where they metamorphose into macroschizonts and stimulate host cell growth. Following their maturation into microschizonts and ultimately uninucleated merozoites by merogony discharged into the bloodstream, macroschizonts present opportunities for merozoite multiplication and development. inside of RBCs, piroplasm forms. This stage produces the illness and has no effect on ticks (6). Theileria's normal life cycle includes the secretion of sporozoites. The parasite undergoes tick gut syngamy to form a zygote after sporozoites invade leukocytes during tick suckling into the feeding site and multiply via merogony. Following this, merozoites are released and enter RBCs to form the piroplasm stage. During the subsequent life cycle feeding, larval or nymph tick vectors ingest piroplasms and liberate the parasite. the only diploid stage, the zygote divides into motile kinetes that infect the tick's gut epithelial cells, move to the haemolymph, and then infect the salivary glands. After tick feeding and initiation, sporogony causes nymphs or adult ticks to multiply sporozoites in salivary gland acini prior to inoculation at the feeding site Figure 1. (7).



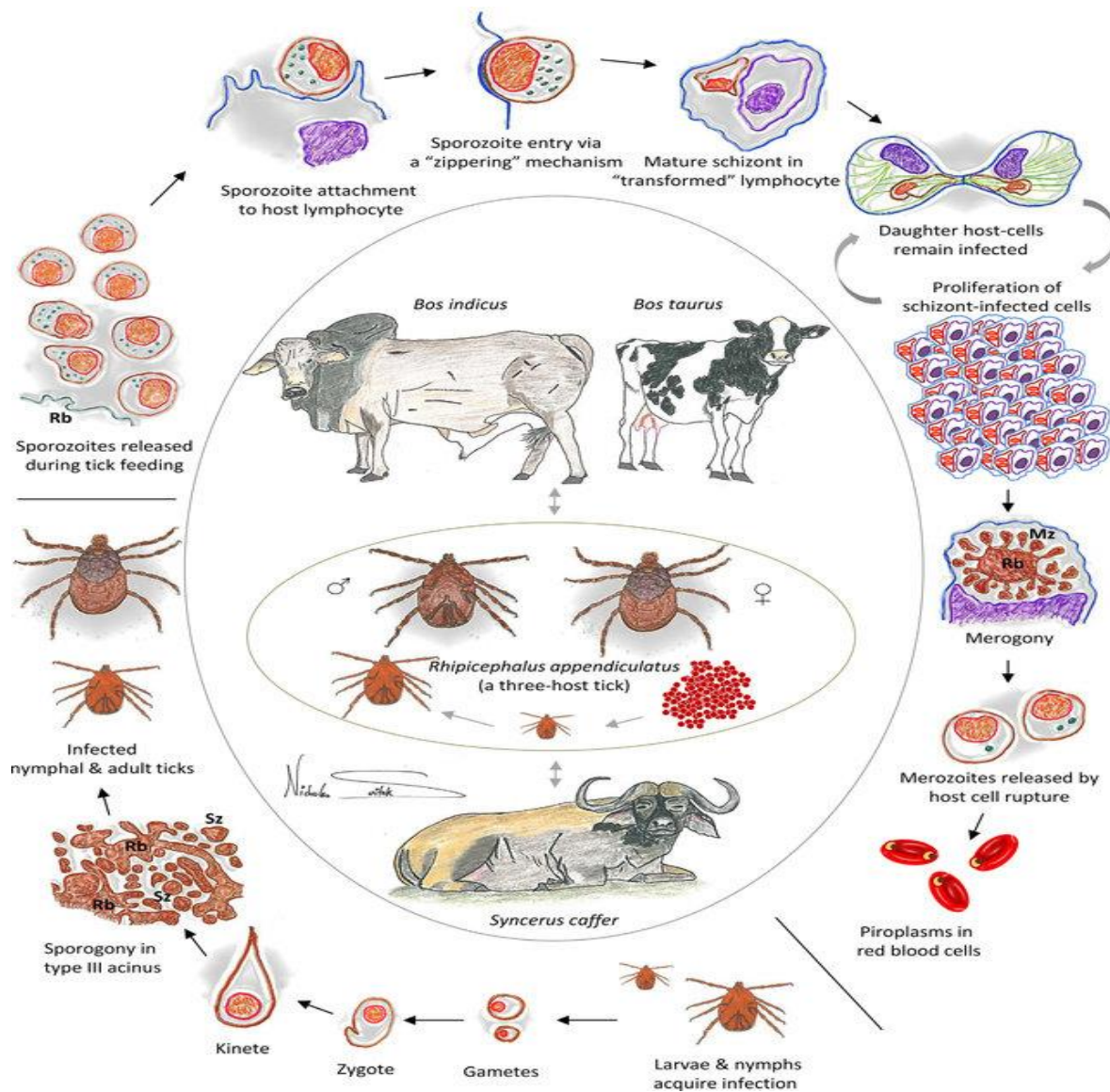


Figure 1: The life cycle of *Theileria* (Bishop et al., 2004).

Transmission of *Theileria* infection

Theileria species only transmits transstadially from one tick stage to the next because ticks are frequently found in vast populations on camels. Ticks in the Ixodidae family are known to carry a variety of infections, ranging from viruses to helminths, which vertebrates can contract. Among these many illnesses, ticks can convey *Theileria* infection. (8). These three tick stages are related to each other because the nymph, larvae, and adult may not all feed on the same host. The infection infects the nymph and larval instars of the tick through a blood meal, and they then move on to the next stage by attaching to a new host. When a nymph or larva is fed, the tick contracts an infection, which spreads to the tick's subsequent stage of life. Adults are more efficient vectors than nymphs. (9). Given that female ticks of the species *Hyalomma* have more type III acini in their salivary glands than males do, female ticks are better at spreading disease. Furthermore, in order to inhibit tick attachment responses, female ticks possess two



histamine-binding proteins. *H. dromedarii* and *A. lepidum* are the names given to two species of female ticks that were collected from Egyptian slaughterhouse camels. The most common kind identified in camels with infestations was *H. dromedarii* (10). Ticks were discovered on several areas of the camel's body; in certain instances, they were seen on the eyelids and inside the ears. The camel's tail area had the highest number of tick reports, with the abdomen being the second most severely infected area. Ticks in the genus *Hyalomma* are a suitable companion for both survival and questing behaviors. Numerous significant species of *Hyalomma* have been reported to spread, including *T. annulata* in various regions of the world, *Hyaloma detritum* in North Africa, *H. detritum* and *H. excavatum* in the former Soviet states, and *H. truncatum* in Africa, in Central Asia the *H. dromedarii*; and in India *H. marginatum*, *H. anatolicum* (11).

Pathogenesis

There are two categories of schizontal organisms in the pathogenesis of *Theileria* spp.: "transforming" and "non-transforming." The unchanging *Babesia* because *Theileria* causes anemia due to piroplasm, it is regarded as contagious. The severity of the disease depends on the virulence, causative strain, host's health, age, infection quantity and susceptibility status. Pathogenesis of numerous forms of *Theileriosis* depends on the manufacture of lymphocyte schizonts and piroplasms in RBCs. The parasite reproduces severe lymphocytopenia, jaundice and anemia in both lymphocyte and erythrocyte form (*T. camolensis* produces various schizonts and piroplasms and all infected camels noticed a clinical feature that could indicate the high pathogenicity (12). While *T. mutans*, *T. ovis*, and *T. buffoli* rarely produce schizonts, they can induce varying degrees of anemia in red blood cells when there is a high concentration of piroplasms. Other *Theileria* species, such as *T. annulata*, *T. hirci*, and *T. parva*, are very pathogenic and develop numerous schizonts and piroplasms. Regarding *Theileria annulata* infection, cells infected with this species express significant quantities of mRNA for cytokines, specifically IFN- γ , because the schizont stage is thought to be the most dangerous parasite stage (13). The clinical manifestations and intensity of the infection are closely associated with the degree of leukopenia halted from these cells' maturation in the bone marrow by the parasite's toxic effect, which also increases the number of infected lymphocytes. *Theileriosis* is caused by macro-schizont and affects lymphocytes as well as reticular endothelial disease (14). The pathophysiology of eosinophilia and lymphopenia has been reported; both conditions are suggestive of lymphadenopathy influenced by *Theileria*, a parasite that first multiplies in lymphocytopenia in lymphoid tissue (15).

Clinical signs

After the infected ticks cling to the victim, the incubation period varies from 4 to 14 days. In acute form, the illness may last as short as three to four days, or it may linger for almost twenty days. The disease's occurrence varies based on the type of parasite, the host's susceptibility to the sporozoite inoculum, and the degree of infection directly relates to the sporozoites' initial injection of the inoculum (16). Depending on the parasite-host connection, the infection might progress from peracute to acute to subacute to chronic. *Theileria* infection in camels was characterized by fever, severe emaciation, ocular discharge, sporadic episodes of diarrhea, and



systemic symptoms such as enlargement of the superficial lymph node. Anorexia, infertility, fast deterioration of health, and abortion.(17). The camels were reported to have been infected with *T. camelensis*, which was characterized by pale mucous membrane, hindlimb weakness, higher heart and breathing rates, and constriction of the rumen. *T. annulata* infection, also called tropical theileriosis, is characterized by fever, swollen lymph nodes, conjunctival petechiae, weight loss, exhaustion, diarrhea, and dysentery.. These symptoms are also associated with later stages of the disease. Animals infected with the pre-acute form of the disease can die within three to four days of the first symptom, while the chronic form can persist for one to two months before the animal recovers (18).

Diagnosis of Thoiloriosis

thoiloriosis is diagnosed based on clinical findings, postmortem observations, and vocal tract distribution. Identification of *Thoiloria* spp. via genetic, serological, and microscopic techniques (19).

Treatment

Many theilericidal, including halofuginonolactate, have been used to treat this disease. *Theileria* is an intraerythrocytic parasite that is difficult to treat and for which there is currently no medication that can eradicate infection. Animal recovery is typically life-threatening. The main medication that is active against schizonts is parvaquone (parvexon ND, Bimeda); the recommended injection dose is 20 mg/kg/IM. Buparvaquone, on the other hand, is active against both piroplasms and schizonts; the recommended injection dose is 2.5 mg/kg/IM. Compared to parvaquone, the efficacy is expected to be 92% after a single injection.(20)

Control

The greatest way to prevent Theileriosis effectively is to combine immunization with tick vector management; in clinical instances, chemotherapy is the only treatment option. there is an urgent need for enhanced control tactics given the disease's continued prevalence in endemic countries (21). Chemical groups that include formamidine, carbamate, organophosphate, and synthetic pyrethroid groups are used to classify acaricides. These substances are applied by hand spraying, in dips, or as spray residues. The terms "pour on" and "spot on" were added more recently (22).

References

- 1.Hassan, M.I.; Gabr, H.S.M.; Abdel-Shafy, S.O.B.H.; Hammad, K.M. and Mokhtar, M.M. (2017): Prevalence of tick-vectors of *Theileria annulata* infesting the onehumped camels in Giza, Egypt. *Journal of Egyptian. Society. Parasitology*, 47(2): 425-432
- 2.Zakian, A.; Nouri, M.; Barati, F.; Kahroba, H.; Jolodar, A.; and Rashidi F. (2014). Vertical transmission of *Theileria lestoquardi* in sheep. *Vet Parasitol*, 203: 322- 325.
- 3.Shahnawaz, S.; Ali, M.; Aslam, M. A.; Fatima, R.; Chaudhry, Z. I.; Hassan, M. U. and Iqbal, F. (2011). A study on the prevalence of tick-transmitted pathogen, *Theileria annulata*, and hematological profile of cattle from southern Punjab(Pakistan). *Parasitology research* ,109(4),1155-1160.



- 4.Hassan, A.H.; Salmo, N.A., and Ahmed, S. (2012): Pathological and molecular diagnostic study of Theileriosis in cattle in Sulaimaniyah Province, Iraq. *The Iraqi journal of veterinary medicine*,36(special issue (2)), 306–314.
- 5.Aktas, M.; Sevgili, M.; Dumanli, N.; Karaer, Z. and Cakmak, A. (2001). Elazig Malatya ve Tunceli illerinde tropikal Theileriosis in seroprevalans. *Turk. J. Veter. Anim. Sci.*, 25: 359–363.
6. Aktas, M.; Altay, K.; Ozubek, S. and Dumanli, N. (2012): A survey of ixodid ticks feeding on cattle and prevalence of tick-borne pathogens in the Black Sea region of Turkey. *Vet. Parasitol.* 187: (3-4); 567-571.
- 7.Hassan, A.H.; Salmo, N.A., and Ahmed, S. (2012): Pathological and molecular diagnostic study of Theileriosis in cattle in Sulaimaniyah Province, Iraq. *The Iraqi journal of veterinary medicine*,36(special issue (2)), 306–314
- 8.Ibrahim, A. M.; Kadle, A. A. H. and Nyingilili, H.S. (2017): Microscopic and Molecular Detection of Camel Piroplasmiasis in Gadarif State, Sudan. *Hindawi Publishing Corporation Veterinary Medicine International Volume*, <http://dx.doi.org/10.1155/2017/9345231>.
- 9.Forsyth, L.M.; Minns, F.C.; Kirvar, E.; Adamson, R.E.; Hall, F.R.; McOrist, S.; Brown, C.G. and Preston, P.M. (1999): Tissue damage in cattle infected with *Theileria annulata* accompanied by metastasis of cytokine-producing, schizontinfected mononuclear phagocytes. *J. Comp. Pathol.*; 120: 39-57.
- 10.Geysen, D.; Bishop, R.; Skilton, R.; Dolan, T. T. and Morzaria, S. (1999): Molecular epidemiology of *Theileria parva* in the field. *Tropical Medicine and International Health* 4, A21-A27.
- 11.Hamidinejat, H.; Razi Jalali, M. and Noori, M. (2008): Report on Clinical Theileriosis in a One-year Old Camel in Khuzestan Province, the Fifteenth Iranian Veterinary Congress.
12. Gharbi, M. and Darghouth, M. A. (2015): Control of tropical Theileriosis (*Theileria annulata* infection in cattle) in North Africa. *Asian Pacific Journal of Tropical Disease*, 5(7), 505- 510
- 13.El Hag, L. M. and Salih, D. A. (2015): Prevalence of Tropical Theileriosis in Nyala Dairy Farms, South Darfur State, Sudan. *Sudan J. V. Res.*, 30: 7-12.
- 14.Islam, M. K.; Alim, M. A.; Tsuji, N. and Mondal, M. M. (2006): An investigation into the distribution, host-preference and population density of ixodid ticks affecting domestic animals in Bangladesh. *Trop Anim Health Prod* 38: 485-490.
- 15.Joshua, K.; Turaki, A. U.; Egwu, G. O.; Mani, A. U.; Saidu, M. K.; Abdullahi, J. G. and Kumshe, H. A. (2008): Haemoparasites of camels (*Camelus dromedarius*) in Maiduguri, Nigeria. *Animal Research International*, 5(2):838-839.
16. Junlong, L.; Li, Y.; Liu, A.; Guan, G.; Xie, J.; Yin, H. and Luo, J. (2015): Development of a multiplex PCR assay for detection and discrimination of *Theileria annulata* and *Theileria sergenti* in cattle. *Parasitology research*, 114(7), 2715-2721.
- 17.Kaaya, G. P. (2000): The potential for anti-tick plants as component of an intergrated tick control stratege. *Tropical Veterinary Diseases* 916 pp:576-582.
- 18.Hamed, M.I.; Zaitoun, A.M.A.; El-Allawy, T.A.A. and Mourad, M.I. (2011). Investigation of *Theileria camelensis* in camel infested by *Hylomma dromedarii* in upper Egypt. *Journal of advance veterinarresearch*,1,4-7.



-
19. Abd-Elmaleck, B.S.; Abed, G.H. and Mandourt, A.M. (2014): Some Protozoan Parasites Infecting Blood of Camels (*Camelus dromedarius*) at Assiut Locality, Upper Egypt. *J. Bacteriol. Parasitol.*, 5: 18
20. Ismael, A.; Swelum, A.; Khalaf, A. and Abouheif, M. (2014): Clinical, Haematological and Biochemical Alterations Associated with an Outbreak of Theileriosis in Dromedaries (*Camelus dromedarius*) in Saudi Arabia. *Pak. Vet. J.* 34(2), 209- 213.
21. Kamani, J.; Usman, T.A.; Onyemaechi, E.G.; Usman, M.A. and Kida, S. M. (2008): Hemoparasite OF Camels (*Camelus dromedarius*) in Maiduguri, Nigeria, *Anim Res Int* 5:838-839.
22. Abdelhaleem, A. A. (2018): Prevalence and Risk Factors of camel theileriosis in Northern State, Sudan. Sudan University of Science and Technology College of Graduate Studies.