

PREVENTION AND TREATMENT OF SHEEP POISONING BY HELIOTROP PLANT

G. T. Abduraimova
Basic Doctoral Student

B. A. Elmurodov
Professor, Scientific Supervisor
Veterinary Scientific Research Institute

Abstract:

This article was prepared on the basis of literature data the article contains information about heliotrope plant species and livestock poisoning from heliotrope.

Keywords: heliotrope, poisoning, hairy heliotrope, elliptic heliotrope, olga heliotrope, prostrate heliotrope, alkaloid.

Introduction

In recent years, a number of decisions have been made by our government in order to develop livestock breeding in our country, ensure food safety, and meet the demand for livestock products (meat, milk, eggs). In particular, the decision of the President of the Republic of Uzbekistan Sh.M. Mirziyoyev of June 1, 2017 on "Measures for organizing the activities of the State Veterinary Committee of the Republic of Uzbekistan" No. on measures" PF-5696, decision PQ-4254 dated March 28, 2019 "On organizing the activities of the State Committee for Veterinary and Animal Husbandry Development of the Republic of Uzbekistan" in order to ensure the rapid development of animal husbandry, aimed at satisfying the daily growing demand of our people for livestock products. However, due to the rapid development of the field, the heliotrope plant, which is found among livestock, causes great economic damage as a result of the death of sheep and lambs [1, 2].

Heliotrope - blue weed, also called common or wild heliotrope. It was discovered in Australia in the 19th century and has been found growing everywhere except the Northern Territory.

Heliotrope is an annual herb for summer planting, growing in meadows or open, cultivated fields. The stem is branched, grows up to 300 mm in height, covered with coarse white hairs. The leaves are gray-green in color, oval in shape, arranged alternately on the stem and have small white flowers.

The seeds germinate in warm, moist conditions after each late spring or summer rain, and seedlings grow where there is no shade and competition from perennials. It is more common in wheat-growing areas than in pastures, and it causes problems during harvest. Heliotrope has a very high seed potential and the seeds can survive for many years.



This plant contains poisons or toxins known as pyrrolizidine alkaloids. Toxins are present at all stages of growth and in all parts of the plant, including the seed [6].

Hairy heliotrope (*Heliotropium dasyocarpum* Ldb.). The stem is shoulder-branched, 20-50 cm high, the leaves are ovate or oblong, branch-shaped, the upper part is tufted, the flowers are whitened at the ends of the stems and branches, there are four nut fruits.

Elliptical heliotrope (*N. ellipticum* Ldb.). The height of the stem is 10-40 cm, the flowers are ovate-oblong-oval leaves on hairy long petioles, and there are four nut fruits. Widespread in the Caucasus, Central Asia. It often grows as a weed in semi-deserts and semi-steppes.



Heliotrope hairy (*N. ellipticum* var. *lasiocarpum* M. Pop., *N. lasiocarpum* Fisch. et Mey.). The stem

is branched, 20-50 cm high, the leaves are oval-elliptic or almost round, the flowers are located on one side of the flowering branch, the leaves are small at the tops of the stems and branches, the fruit is small, tetrahedral, when ripe it is divided into four hairy nuts.

It is distributed in the Republics of Central Asia, in the sand dunes of the southeast. Fields planted with wheat and barley are affected by heliotrope, it also grows in gardens, steppes, along roads.

Olga heliotrope (*N. Olga* Bge.). The stem is 10-40 cm high, branched, downy hairy, the leaves are ovate or rounded ovate, single or double, one-sided, 5 cm long fruit - ovoid, bare nuts. It has been reported to meet in the countries of Central Asia [2,3].

Prostrate heliotrope (*N. supinum* L.). Annual plant. The stem is branched, elevated, 5-20 cm long, white-gray-fluffy leaves are ovate or oblong, hairy flowers are white in color, one persistent fruit nut is found in lateral or terminal folds.

Heliotrope poisoning (heliotropotoxicosis). Heliotropes are alkaloid plants. Heliotrin C₁₆H₂₇NO₅, liziocarpine C₂₁H₃₃NO₇ (GP Menshikov, 1932) found in elliptical (pubescent) heliotrope; in prostrate heliotropia, supine C₁₅H₂₅NO₄ alkaloid (G.P. Menshikov, E.L. Gurevich, 1949). Alkaloids are also found in hairy heliotropes with a fragrant smell (A. A.



Ataev, 1958) [4,5].

The amount of heliotrim in a dried plant is 0.3 - 0.4%, liziocarpin - 0.03%; the amount of alkaloids in the seeds is more significant (D.N.Sahibov, 1957).

Heliotrope pubescent (*Heliotropium lasiocarpum*) is a disease that develops as a result of eating cereal products mixed with seeds.

The disease was first recorded in the 30s of the 20th century. In Central Asia (sheep mortality



was observed up to 20-30%). Poisoning has also been reported as a result of eating bread and porridge made from grains mixed with heliotrope seeds.

In 1945, research conducted by H.H. Zener showed that grain used in the feed of diseased animals contained a mixture of heliotrope seeds. The ground part (and seeds) of the plant contain heliothrine and lasiocarpine alkaloids, discovered by G. P. Menshikov in 1932-1934, which seriously damage the liver parenchyma and vessels of the animal. Liver damage is associated with decreased protein synthesis, increased hepatic breakdown, decreased glycogen, and concomitant increased mobilization of fat from hepatic depots, followed by the development of moderate fatty liver. It is characterized by a decrease in lipolytic enzymes and gas exchange in the liver.

Etiology. Intoxication is observed in pigs, less fattened cattle, sheep and chickens, as a result of feeding with grain feed containing up to 5% and more heliotrope seeds. Poisoning occurs as a result of eating the vegetative parts of the plant in the budding phase of the black sheep in the pasture.

Pathogenesis. The onset of heliotrope poisoning is the alkaloids heliothrine and lysiocarpine, which are present in the seeds from 0.02% to 3% or more, and to a lesser extent in the vegetative parts of the plant, which are mainly neurohepatotropic.

Clinical signs. At the beginning of the disease in cattle, refusal of food, general weakness, lethargy, yellowing of mucous membranes, clay-colored faecal masses are noted. On the 8-9th day of the disease, the weakness of the heart is accompanied by swelling of the subcutaneous tissue. Visible mucous membranes are hyperemic, with bleeding. At the beginning of the disease, pigs are lethargic, lie down for a long time, have poor or no appetite, their feces are dark in color and have an unpleasant smell.

In the acute period of the disease, depression increases, reaction to the environment is weakened, pain sensitivity decreases, vomiting is observed, and the animal falls down when trying to get up. From time to time, contraction of individual muscles of the body is noted. Animals die with continuous clonic-tonic convulsions, swimming movements of the legs.

At the beginning of intoxication in sheep, salivation, vomiting and pain in the liver are observed. On the 5-7th day of intoxication, painful urination occurs, urine mixed with blood.

In acute intoxication of birds, there is a lot of bleeding in the ears, skin of the abdomen and under the mucous membranes. Feces are liquid, with a mixture of blood. Death occurs on the 2-10th day of the disease.

In chronic intoxication, general weakness, abdominal distension, fatigue and death are noted.

Pathanatomical changes. Rigor mortis is soft, noncoagulable, and visible mucous membranes are pale and icteric. Pale yellow, lemon-yellow fluid with an unpleasant odor appears in the abdominal cavity.

Muscles are light yellow or strong yellow. The liver is enlarged, plethoric, yellow or yellow-brown in color, chronically shrunken and dense to the touch (atrophic cirrhosis). In the acute period of poisoning in pigs, ulcers are found on the mucous membranes of the small intestine.

Diagnosis. It is based on characteristic clinical signs, anamnesis data (hepatitis with ascites) and the results of laboratory analysis of food.



Treatment. Feed and pasture should be changed. Poisoned animals are injected intravenously with 150-200 ml of 40% glucose solution for large animals, 100-150 ml for small animals, as well as 75-100 ml of 10% solution of hexamethylenetetramine, 500-1000 ml of hemodesis, repeated. 10-12 hours 15-30 ml of campolon or 10-15 ml of hepalon are injected into the muscle. The use of vitamins - retinol, tocopherol and calciferol is indicated.

Prevention. In order to eliminate heliotrope, weeding is carried out twice before flowering and fruiting of cultivated fields, especially grain crops with spikes, and deep plowing in autumn [3,4,6].

References

1. Ibadullayev F.I., "Qishloq xo'jalik hayvonlarining patologik anatomiyasi" Toshkent "O'zbekiston" 2000 y.
2. Ibadullayev F.I., Elmurodov B., Abdusattorov A. "Qo'zilarida uchiraydigan aralash kasalliklar" Vet. J 1997-№1 18-19 bet
3. Scott Ison, Albury and Helen Peam, Elizabeth McArthur. Secondary photosensitisation in lambs due to crystal associated cholangiohepatopathy while grazing common heliotrope (*heliotropium europaeum*) probably caused by panicum sp. Toxicity Posted Flock & Herd September 2015
4. Button, C., Paynter, D.I., Shiel, M.J., Corlson, A.R., Paterson, P.J., Lyford, R.L. Crystal-associated cholangiohepatopathy and photosensitization in lambs. Aust. Vet. J 1987; 64:176-180
5. Cunningham GM, Mulham WE, Milthorpe PL & Leigh JH. Plants of Western New South Wales. CSIRO Publishing 1981
6. Hunt JR. The ecology of common heliotrope (*Heliotropium europaeum* L.) in a Mediterranean dry-land cropping system. PhD thesis, Faculty of Land and Food Resources, The University of Melbourne 2005
7. Elmurodov, B. A. (2002). Detection of mixed bacterial infections in calves. *Journal of Agriculture of Uzbekistan. Tashkent*, 3, 63.
8. Эльмуродов, Б. А. (2003). Смешанные инфекции телят. *Ветеринарная патология*, (2), 52-53.
9. Azamov, V., Elmurodov, B., Parmanov, J., & Abdalimov, S. (2004). Changes in the intestinal system in colibacillosis. In *Proceedings of the Third Republican Scientific-Practical Conference, Samarkand* (pp. 9-12).
10. Abdalimov, S. A., Parmanov, J. M., & Elmurodov, B. A. (2004). Sheep pasteurellosis//Third Res. II-Amal. konf. ma'r. Collection of texts.
11. Элмуродов, Б. А. (2005). Клинические изменения при смешанных бактериальных инфекциях птиц.
12. G'aniyev, I., & Elmurodov, B. A. (2008). Course and clinical signs of sheep pasteurellosis. In *Four. ilm.-amal. konf. ma'r. text collection. Samarkand* (pp. 94-96).
13. Duskulov, V. M., Elmurodov, B., & Meyliev, M. (2018). Highly profitable sector of beekeeping. *Veterinary Medicine*, 12.



14. Элмуродов, Б. А., Турдиев, А. К., & Набиева, Н. Куёнчилик укув кўлланма. *Самарканд-2018*, 72-73.
15. Эльмурадов, Б. А., Наврузов, Н., & Курбонов, Ф. (2019). Патологоанатомические изменения при смешанных бактериальных инфекциях птиц.
16. Эльмурадов, А., & Эльмурадов, Б. А. (2019). Содержание нуклеиновых кислот в стенках двенадцатиперстной кишки у каракульских овец различного возраста и в разные сезоны года.
17. Navruzov, N. I. The Role of Immunostimulants in the Prevention of Colibacillosis, Salmonellosis and Pasteurellosis in Calves. *International Journal on Integrated Education*, 3(8), 232-234.
18. Элмуродов, Б. А., & Эшбуриев, С. Б. (2021). ТОВУҚЛАРДА МИНЕРАЛЛАР АЛМАШИНУВИ БУЗИЛИШЛАРИНИНГ КЛИНИК БЕЛГИЛАРИ. *ВЕСТНИК ВЕТЕРИНАРИИ И ЖИВОТНОВОДСТВА*, 1(1).
19. Муродов, Х., Элмуродов, Б., Шодиева, У., & Ахмедов, Б. (2021). Профилактика и лечение инфекционного ларинготрахеита птиц. *in Library*, 21(2).
20. Navruzov, N. I., Elmurodov, B. A., & Mamadullaev, G. K. (2021). THE ROLE OF CHITOSAN IN THE PATHOMORPHOLOGY AND IMMUNOPROPHYLAXIS OF COLIBACILLOSIS OF CALVES.
21. Ахмадалиева, Л. Х., Элмуродов, Б. А., & Орипов, А. О. (2021). ПРАВОВАЯ ОХРАНА ЗДОРОВЬЯ ЖИВОТНЫХ И ЭКОСИСТЕМ В НИИ ВЕТЕРИНАРИИ. *ББК 40.0 П78*, 378.
22. Nabieva, N. A., Elmurodov, B. A., & Aktamov, U. B. (2022). Biochemical Changes in Blood in Rabbit Pasteurella's. *Texas Journal of Medical Science*, 13, 115-118.
23. Elmurodov, B. A., Navruzov, N. I., & Kiyamova, Z. N. (2022). Intervention of Bacterial Diseases in Poultry. *INTERNATIONAL JOURNAL OF BIOLOGICAL ENGINEERING AND AGRICULTURE*, 1(4), 8-12.
24. Элмуродов, Б. (2022). Ветеринария илм-фанининг истиқболлари ва соҳани ривожлантиришдаги муҳим вазифалар. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(2), 462-464.
25. Элмуродов, Б., & Исмоилов, У. (2022). Молодняк животных в районах приаралья течение колибактериоза и сальмонеллеза. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(1), 233-235.
26. Элмуродов, Б., & Исмоилов, У. (2022). Текст научной работы на тему Течение колибактериоза и сальмонеллеза молодняка в Приаралье. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(2), 307-309.
27. Эльмурадов, Б. (2022). Перспективы ветеринарии и важные задачи развития отрасли. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(1), 9-12.
28. Sh, N., Elmurodov, B. A., & Eshburiev, S. B. (2022). TUXUM YONALISHDAGI TOVUQLAR MAHSULDORLIGIGA NOVAMIX PREMIKSINING TASIRI. *AGROBIOTEKNOLOGIYA VA VETERINARIYA TIBBIYOTI ILMIY JURNALI*, 476-479.



29. Набиева, Н., Элмуродов, Б., & Сайдуллаев, А. (2022). Эпизотология пастереллиоза кроликов. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(1).
30. Namraqulov, N. S. (2022). SYMPTOMS AND HEMATOLOGICAL INDICATORS OF CALCIUM AND PHOSPHORUS EXCHANGE DISORDERS IN CHICKEN IN EGGLAYING HENS. *Conferencea*, 92-94.
31. Султанова, И., & Элмуродов, Б. (2022). Течение и бактериологическое течение сальмонеллы у кроликов методы проверки. *Перспективы развития ветеринарной науки и её роль в обеспечении пищевой безопасности*, 1(2), 187-191.
32. BA, Elmurodov, et al. "Pathomorphological Changes in Poultry Pasteurelliosis, Pullorosis and Colibacteriosis Diseases." (2023).
33. Aktamovich, E. B., Bakhtiyorovich, E. S., & Shokir, H. N. (2023). Prevention of Calcium Phosphorus Exchange Disorders in Chickens. *Web of Semantic: Universal Journal on Innovative Education*, 2(6), 222-228.
34. Мамадуллаев, Г. Х., Элмуродов, Б. А., Джураев, О. А., Джуракулов, О. К., & Файзиев, У. М. (2023). РИФИЗОСТРЕП–НОВЫЙ КОМБИНИРОВАННЫЙ ПРЕПАРАТ ПРОТИВ МИКОБАКТЕРИЙ ТУБЕРКУЛЁЗА. *Эпизоотология Иммунология Фармакология Санитария*, (2), 52-57.
35. Navruzov, N. I., & Elmurodov, B. A. THE ROLE OF CHITOSAN SUCCINATE IN COLIBACILLOSIS OF CALVES AND THE EFFECT ON THE IMMUNE SYSTEM. *СБОРНИК ТЕЗИСОВ*, 50.
36. Nabieva, N. A., & Profissor, B. E. V. (2023). PATHOGISTOGRAM OF PASTEURILLOSIS OF RABBITS. *European International Journal of Multidisciplinary Research and Management Studies*, 3(01), 92-98.
37. Elmurodov, B. A., Abdalimov, S. H., & SHERALIEVA, I. D. Diseases of young animals Samarkand 2016.
38. Алламуродова, М., Киямова, З., & Элмуродов, Б. А. (2024). ИННОВАЦИОННЫЙ ПОДХОД К ПРОФИЛАКТИКЕ ЗАБОЛЕВАНИЙ ЖИВОТНЫХ. *World scientific research journal*, 25(1), 128-133.
39. Набиева, Н. А., & Элмуродов, Б. А. (2024). ҚУЁНЛАР ПАСТЕРЕЛЛЁЗИНИ ДАВОЛАШДА АНТИБИОТИКЛАР САМАРАДОРЛИГИ. *World scientific research journal*, 25(1), 134-140.
40. Navruzov, N. I., Kiyamova, Z. N., & Elmurodov, B. A. (2024). SALMONELLA PULLORUM GALLINARIUM BILAN ZARARLANGAN JO ‘JALARDA PATOMOFOLOGIK O ‘ZGARISHLAR. *World scientific research journal*, 25(1), 141-151.
41. Элмуродов, А. А., Абдуллаева, Ю. У., & Абдуллаева, С. А. (2023). ЭФФЕКТИВНОСТЬ ВЫРАЩИВАНИЯ СЕМЕННЫХ КЛУБНЕЙ СОРТОВ КАРТОФЕЛЯ IN VITRO В УСЛОВИЯХ ЗЕРАВШАНСКОЙ ДОЛИНЫ. *Бюллетень науки и практики*, 9(1), 173-181.



42. Элмуродов, А. А., & Абдуллаева, Ю. У. ЭФФЕКТИВНОСТЬ ВЫРАЩИВАНИЯ СЕМЕННЫХ КЛУБНЕЙ СОРТОВ КАРТОФЕЛЯ IN VITRO В УСЛОВИЯХ ЗЕРАВШАНСКОЙ ДОЛИНЫ.
43. Элмуродов, Б. А., Наврузов, Н. И., Набиева, Н. А., Ахмадалиева, Л. Х., & Киямова, З. Н. Инновационные вакцины для профилактики пастереллеза кроликов и других животных. In *Современные достижения в решении актуальных проблем агропромышленного комплекса: материалы международной научно-практической конференции, посвященной 100-летию Института экспериментальной ветеринарии им. СН Вышелесского (Минск, 15-16 сентября 2022 г.)* (pp. 282-284).
44. Газнакулов, Т., & Хушназаров, А. (2023). Литературный обзор по истории развития эпизоотологии и изучения бешенства. in *Library*, 1(2), 7-9.
45. Davlatov, R. B., & Khushnazarov, A. K. (2024). Diagnosis and chemoprophylaxis of rabbit eymeriosis. In *E3S Web of Conferences* (Vol. 480, p. 03020). EDP Sciences.
46. Khushnazarov, A. X. (2022). OBZOR LITERATURNYX DANNYX PO KHMIIOTERAPII I KHMIIOPROPHYLAKTIKI EYMEROZA KROLIKOV. *Journal of PEDAGOGS*, 23(2), 83-86.
47. Хушназаров, А. Х. (2022). ОБЗОР ЛИТЕРАТУРНЫХ ДАННЫХ ПО ХИМИОТЕРАПИИ И ХИМИОПРОФИЛАКТИКИ ЭЙМЕРИОЗА КРОЛИКОВ. *PEDAGOGS journali*, 23(2), 83-86.
48. Избасаров, У., Турдиев, А., Дускулов, В., & Хушназаров, А. (2021). Санитарно-гигиеническая оценка полов в животноводческих помещениях в условиях жаркого климата. in *Library*, 21(1), 214-217.
49. Izbasarov, U., Turdiev, A., Duskulov, V., & Khushnazarov, A. (2021). Sanitary and hygienic assessment of floors in livestock buildings in a hot climate. *Library*, 21(1), 214-217.
50. Izbasarov, U., Khushnazarov, A., & Khamroev, A. (2020). Treatment of dermatological and gynecological diseases of humans and animals. *Library*, 20(4), 296-299.
51. Izbasarov, U., Khushnazarov, A., Khamraev, A. K., & Izbasarov, S. U. (2020). Creation of new domestic phyto-tissue preparations for veterinary medicine. *Library*, 20(4), 296-299.
52. Избасаров, У., Хушназаров, А., & Хамроев, А. (2020). Лечение дерматологических и гинекологических заболеваний человека и животных. in *Library*, 20(4), 296-299.
53. Избасаров, У., Хушназаров, А., Хамраев, А. Х., & Избасаров, Ш. У. (2020). Создание новых отечественных фито-тканевых препаратов для ветеринарной медицине. in *Library*, 20(4), 296-299.

